

# Microscopy

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<sup>1</sup>Using material from Davidson and Abramowitz: Optical Microscopy

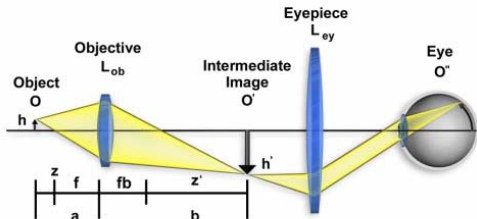
# Microscopy

Optical microscopy – since 17th century; Jensen, van Leeuwenhoek, Galilei, . . .



# Finite-Tube Length Microscope

## Finite-Tube Length Microscope Ray Paths

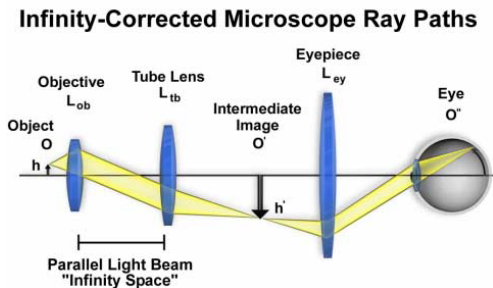


- ▶ magnification of the objective  $\frac{b}{a}$
- ▶ magnification of the eyepiece  $\frac{25 \text{ cm}}{f_{\text{eyepiece}}}$
- ▶ thin-lens equation

$$\frac{1}{a} + \frac{1}{b} = \frac{1}{f}$$

- ▶ narrow range of image distances
- ▶ specifically corrected optical systems with matching eyepieces

# Infinite-Tube Length Microscope

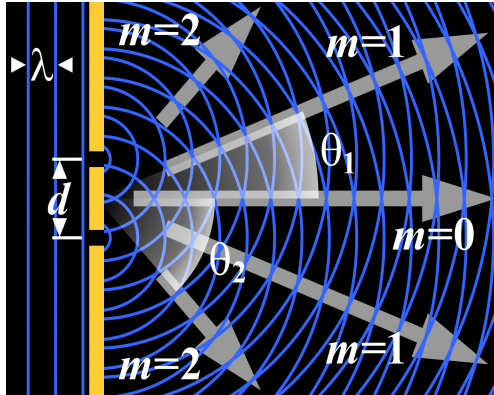


- ▶ Modern design (since 1980s)
- ▶ Objective magnification determined by  $\frac{f_{tb}}{f_{ob}}$
- ▶ Infinity space to add polarizers, prisms, retardation plates. . .
- ▶ Independently changeable objective and eyepiece

# Image Formation

- ▶ Direct/undeviated light
- ▶ Deviated/diffracted light, out of phase
- ▶ Constructive/destructive interference

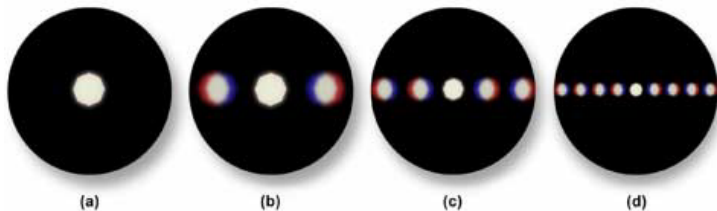
# Diffraction



Position of maxima:

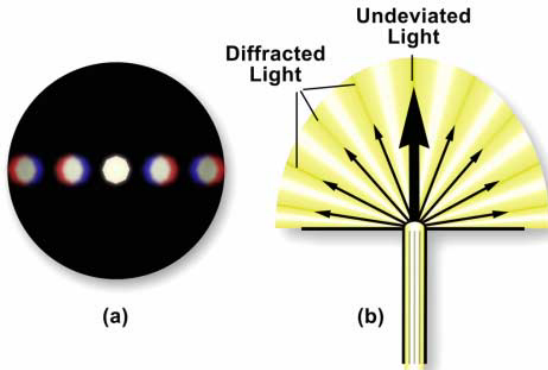
$$d \sin \theta = n\lambda, \quad n \in \mathbb{Z}$$

# Line Grating Diffraction Patterns



- ▶ line phantom
- ▶ close diaphragm
- ▶ telescope, observe the rear focal plane of the objective
- ▶ (a) no phantom, (b)  $10\times$ , (b)  $40\times$  (higher NA), (c)  $60\times$  (highest NA)
- ▶  $0^{\text{th}}$  order,  $1^{\text{st}}$  order image

# Diffraction



- ▶ constructive/desctructive interference
- ▶ specimen = superposition of complex gratings (*Ernst Abbe*)
- ▶ to resolve image, at least 0<sup>th</sup> order and 1<sup>st</sup> order images must be captured
- ▶ more orders captured → better accuracy



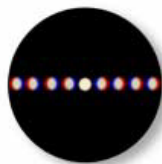
# Slit and Grid Diffraction Patterns



(a)



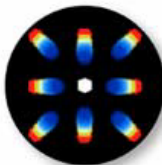
(b)



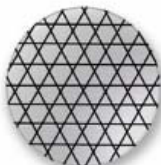
(c)



(d)



(e)



(f)



(g)

# Slit and Grid Diffraction Patterns



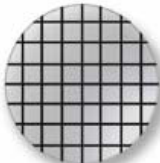
(a)



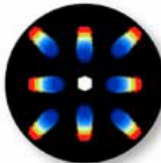
(b)



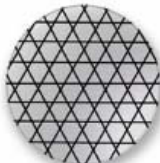
(c)



(d)



(e)



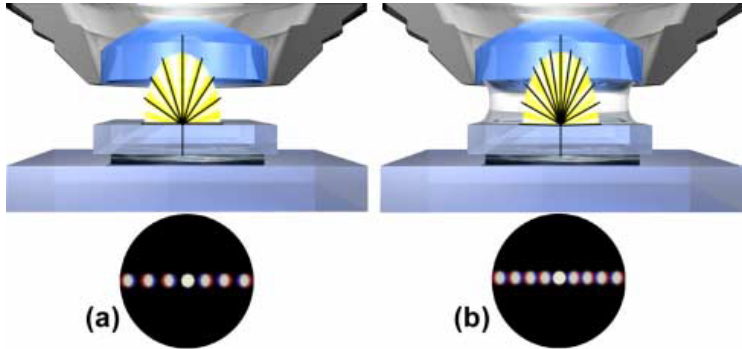
(f)



(g)

- ▶ Diffraction patterns behave like Fourier transforms of the sample
- ▶ Fourier optics

# Immersion optics



- ▶ High refractive-index media (immersion oil) reduce diffraction angle
- ▶ → More orders are captured
- ▶ → Better image

# Resolution limit

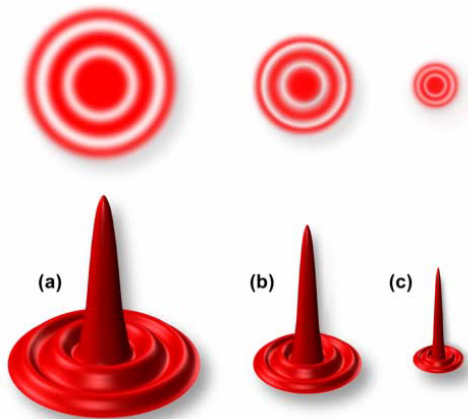
Rayleigh equation:

$$d \approx 1.22 \frac{\lambda}{2 \text{NA}}$$

To improve resolution, use:

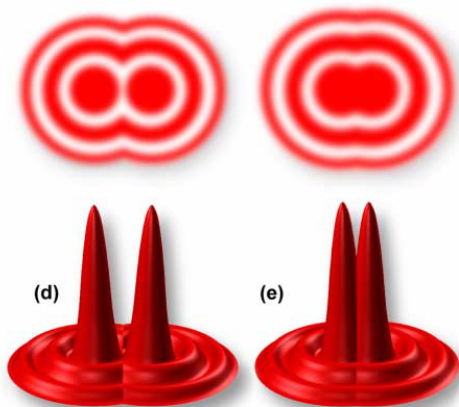
- ▶ Big lenses (big NA)
- ▶ Short wavelength (blue)

# Airy disks



- ▶ NA increases left to right.
- ▶ Impulse response (PSF)

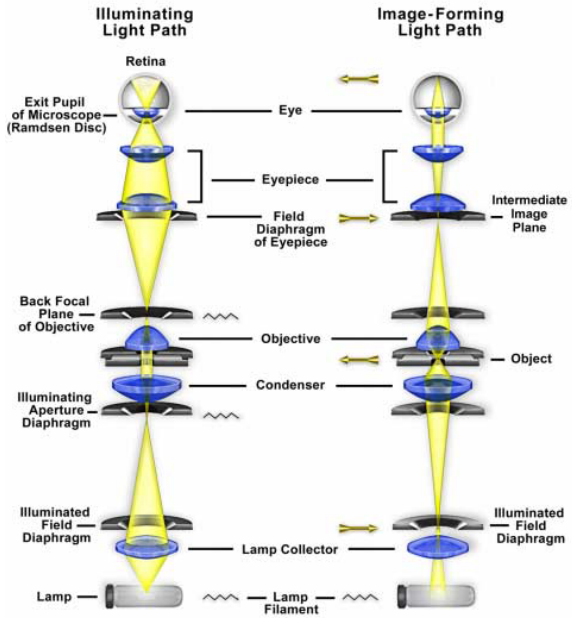
## Airy disks (2)



Resolution limit.

## Köhler illumination

- ▶ Focused lamp image is projected to the diaphragm of a condenser.
- ▶ Field diaphragm controls width of the light bundle.
- ▶ Aperture diaphragm controls the light intensity. Trade-off between diffraction artifacts and glare.
- ▶ Light is not focused on the specimen, illumination is homogeneous.
- ▶ The focal point of image-forming rays is at the level of the specimen.





# Optical Aberrations

- ▶ Geometric aberrations
  - ▶ Spherical — rays on axis and far from the axis do not converge to the same point. Blurred images.
  - ▶ Flat-field — because lenses are curved, the image is curved. Center and off-center not simultaneously in focus.
- ▶ Chromatic aberrations — rays of different color do not converge to the same point

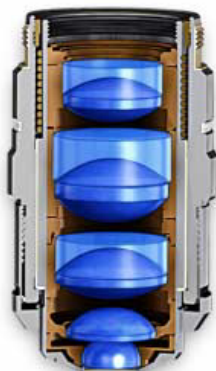
## Optical Correction in Objectives

**Achromatic Objective**



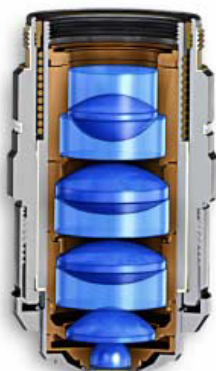
**(a)**

**Fluorite Objective**



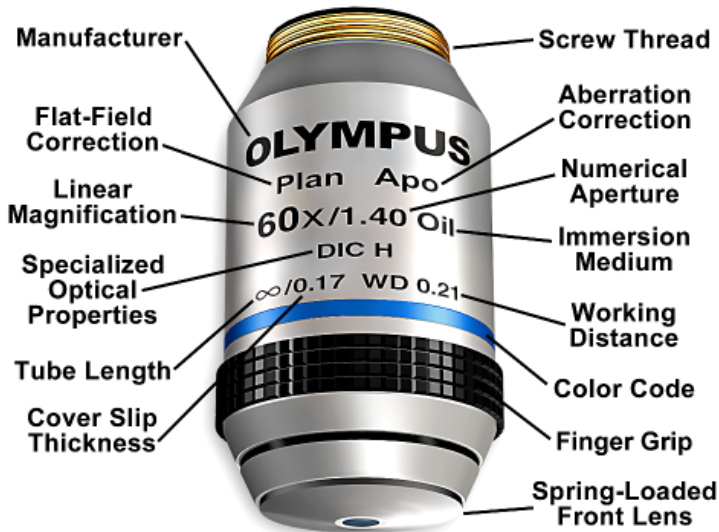
**(b)**

**Apochromatic Objective**

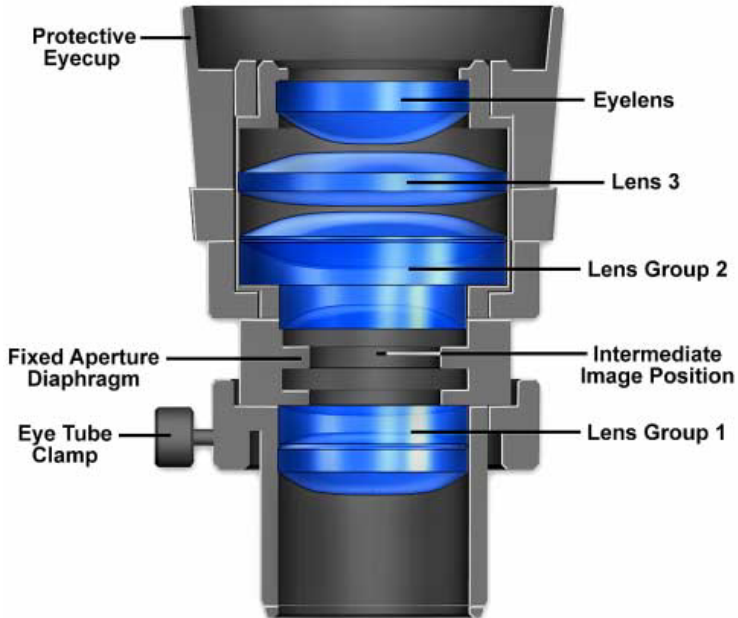


**(c)**

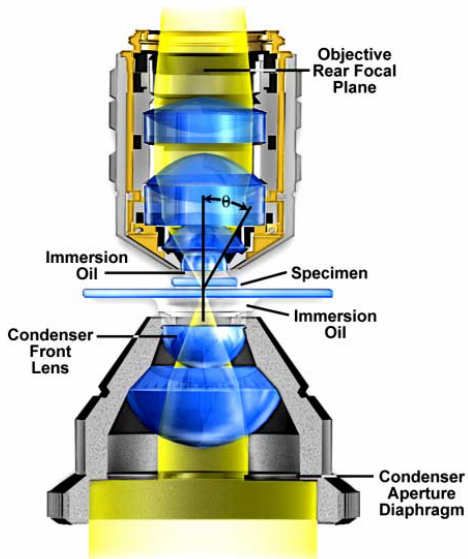
# Objective Specifications



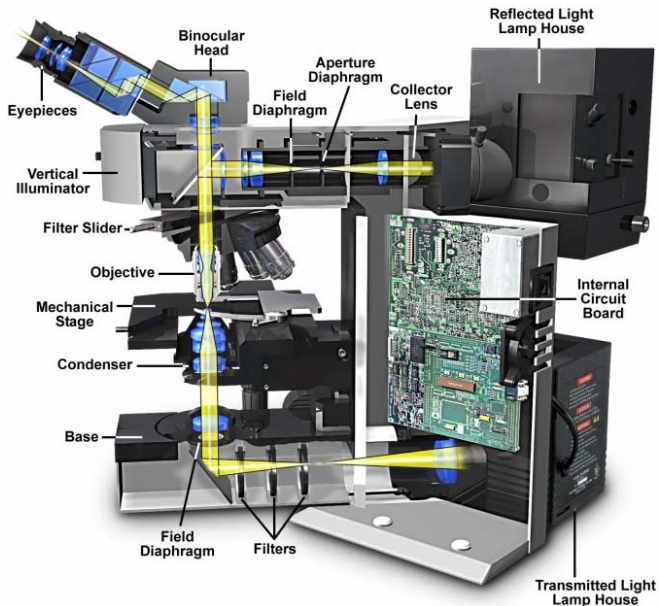
# Eyepiece Cutaway Diagram



# Condenser



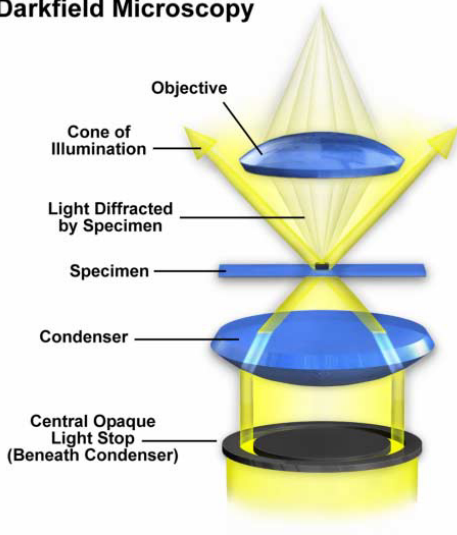
# Reflected light microscope



# Contrast enhancing techniques

- ▶ Dark field microscopy
- ▶ Rheinberg illumination
- ▶ Phase contrast microscopy
- ▶ Polarized light
- ▶ Hoffman modulation
- ▶ Differential interference contrast

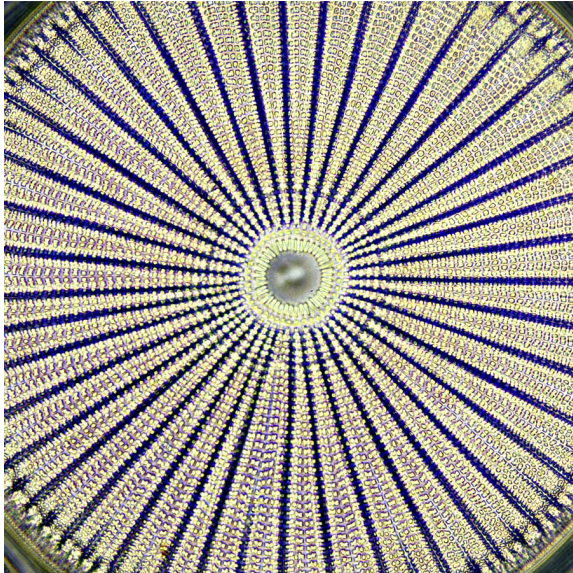
## Darkfield Microscopy



For unstained objects. Appear bright on dark background.

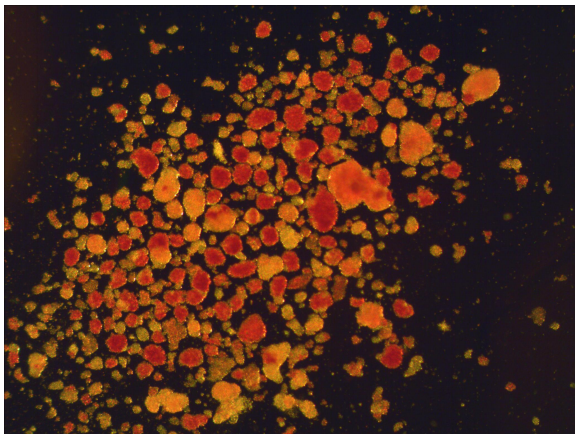


## Darkfield microscopy (2)



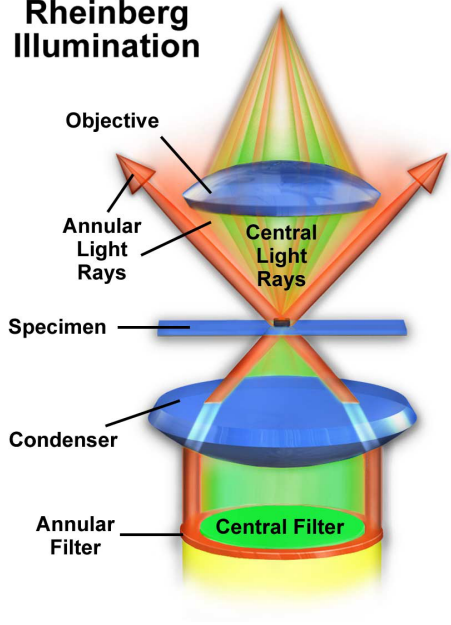
*Arachnoidiscus ehrenbergi*

## Darkfield microscopy (3)



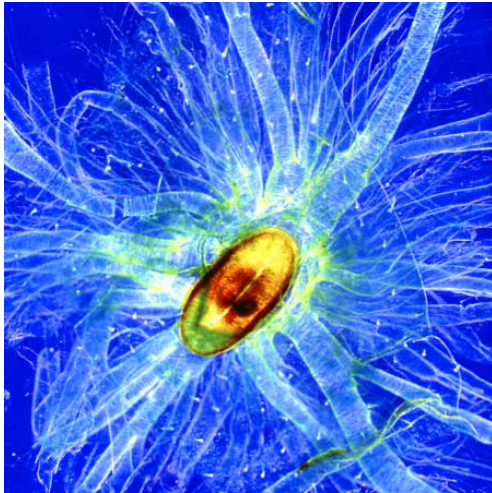
*Langerhans islets*

# Rheinberg Illumination



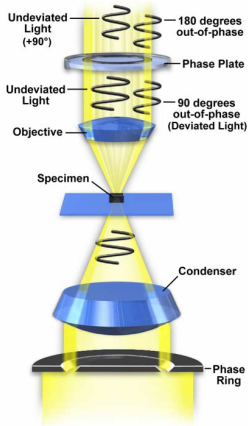
Color annular filters instead of the darkfield stop.

## Rheinberg illumination (2)



silkworm larva

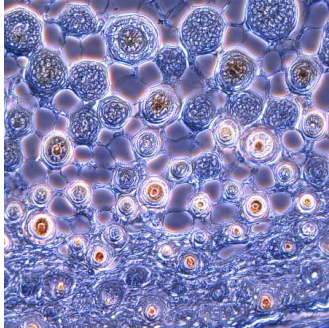
## Phase Contrast Microscopy



Frits Zernike (1930s, Nobel price 1953). Show differences in phase/refractive index.

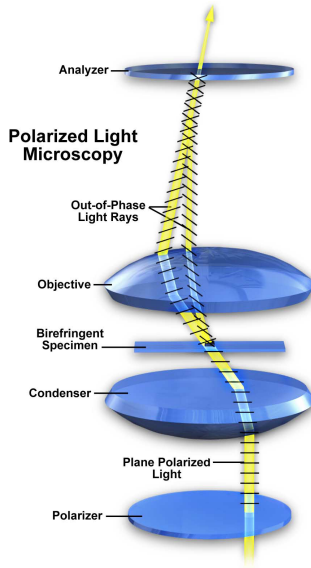
Interference. Slow down/Speed up. direct light → bright/dark contrast

## Phase contrast microscopy (2)



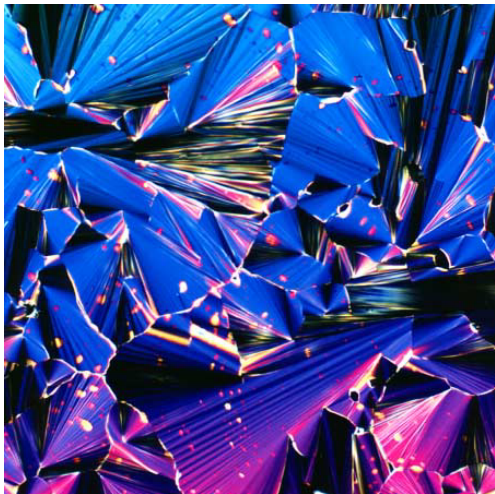
mouse hair cross-section

# Polarized light microscopy



- ▶ different refractive indices for different polarizations

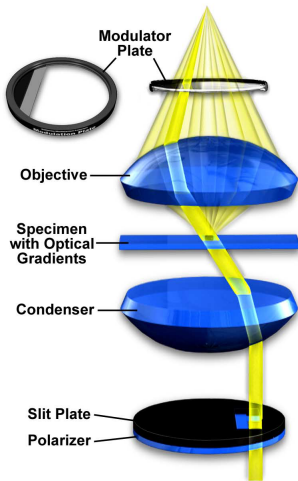
## Polarized light microscopy (2)



DNA

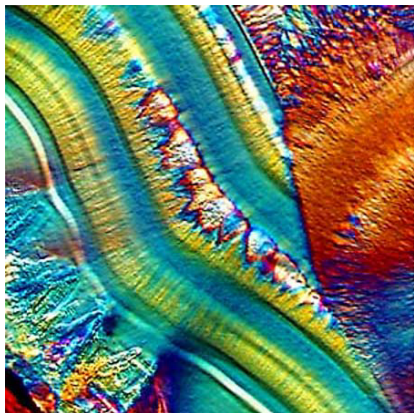


## Hoffman Modulation Contrast Microscopy



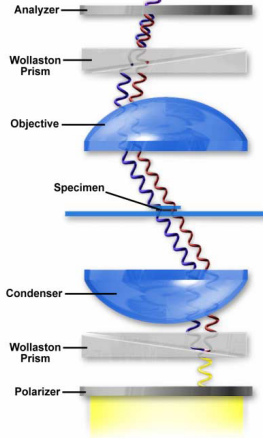
Robert Hoffman (1975). For living and unstained specimens. Detects optical gradients. Image intensity proportional to the derivative of the optical intensity of the specimen.

## Hoffman modulation contrast (2)



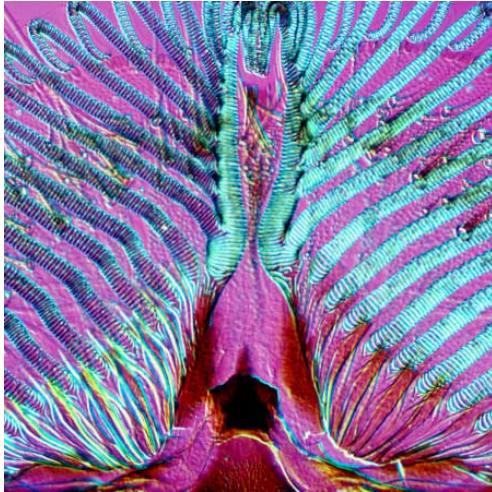
Dinosaur bone

## Differential Interference Contrast Microscopy



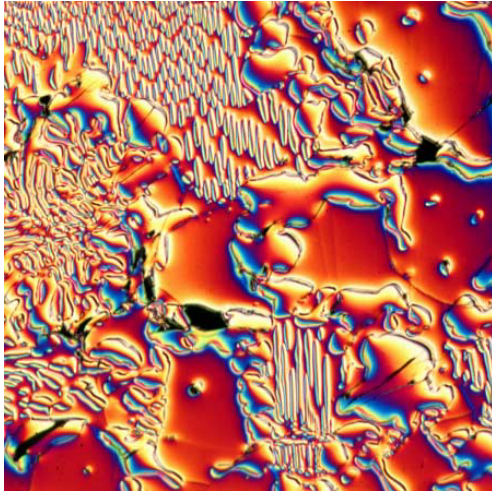
Detects differences in optical paths between two close slightly offset rays (shear).

## Differential interference contrast microscopy (2)



Mouth part of a blowfly.

## Differential interference contrast microscopy (3)

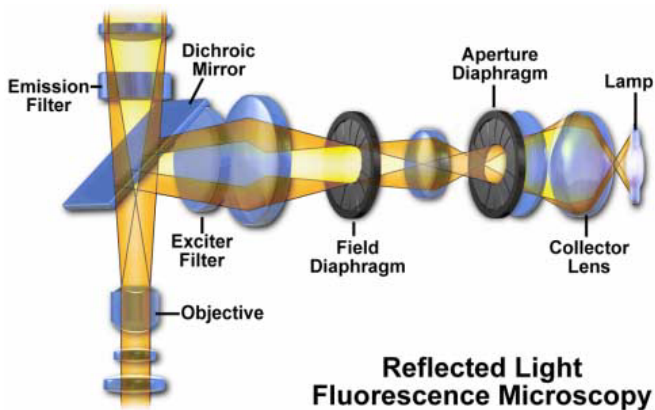


Defects in ferro-silicon alloy.

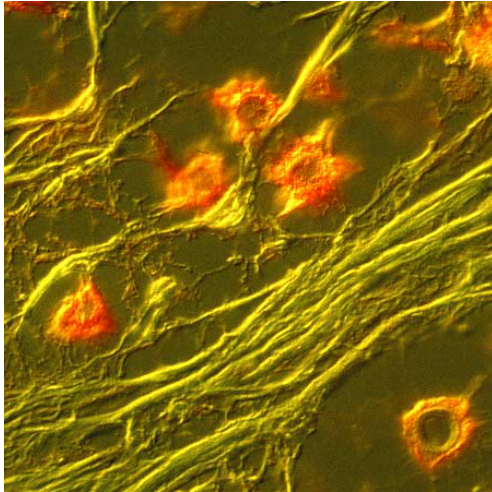
# Fluorescence microscopy

- ▶ fluorescent dyes
- ▶ multiple sensing channels/filters
- ▶ multiple light sources – visible, UV

## Fluorescence microscopy (2)



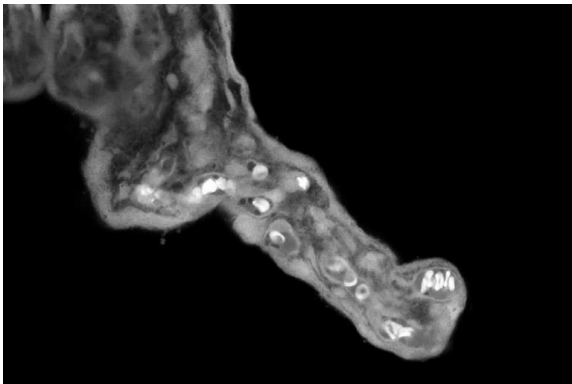
## Fluorescence microscopy (3)



cat brain tissue infected with *Cryptococcus*

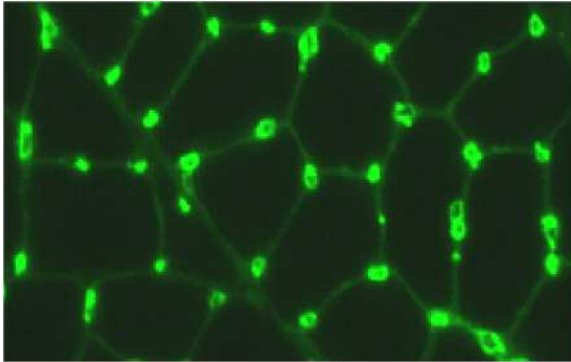


## Other examples images



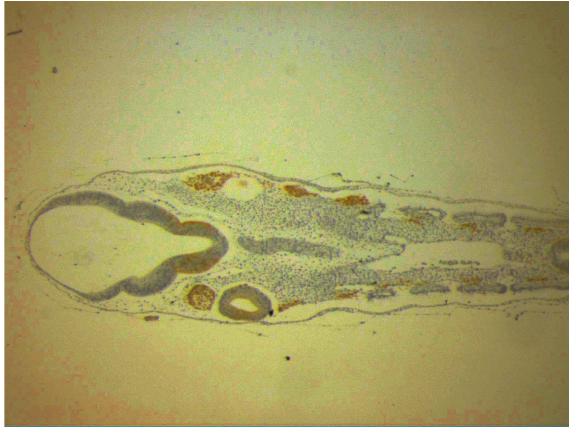
placenta cross-section

## Other examples images



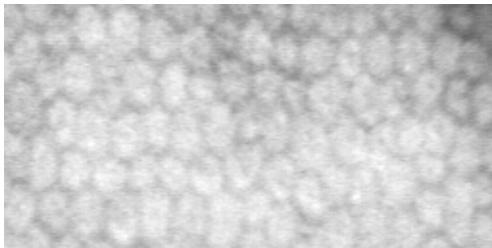
muscle capillaries

## Other examples images



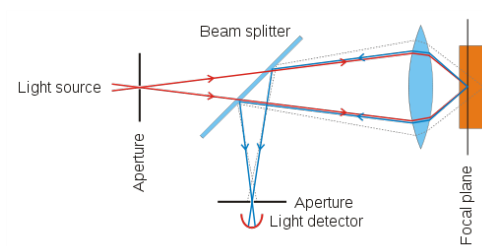
crocodile ear slice

## Other examples images



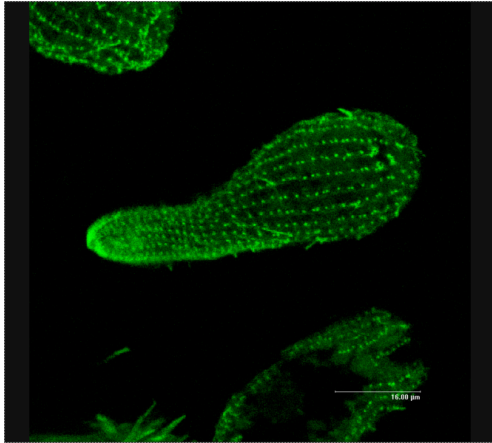
retina

# Confocal microscopy



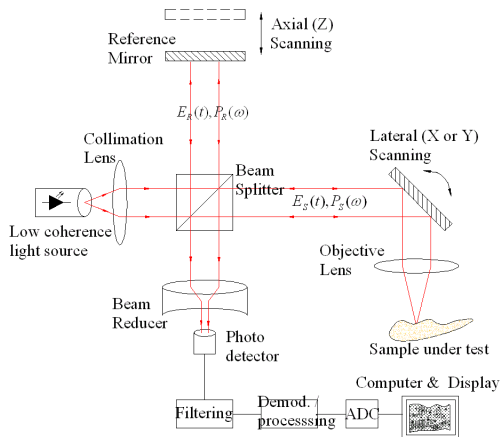
- ▶ Very good resolution
- ▶ Very thin focal plane — 3D imaging
- ▶ Confocal laser scanning
- ▶ Scanning — slow

# Confocal microscopy example



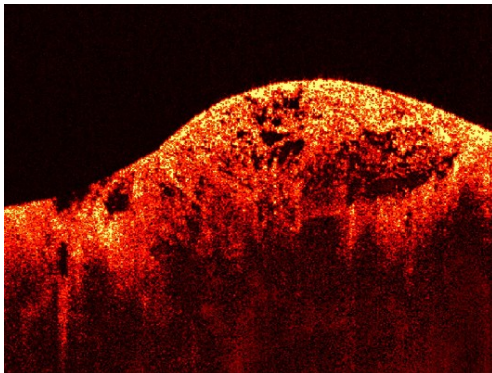
Tetrachimena

# Optical coherence tomography (OCT)



- ▶ 3D imaging
- ▶ Interferometry
- ▶ More penetration than confocal, especially near infrared

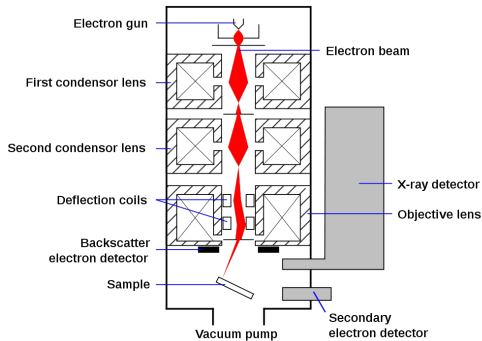
## OCT example



Sarcoma

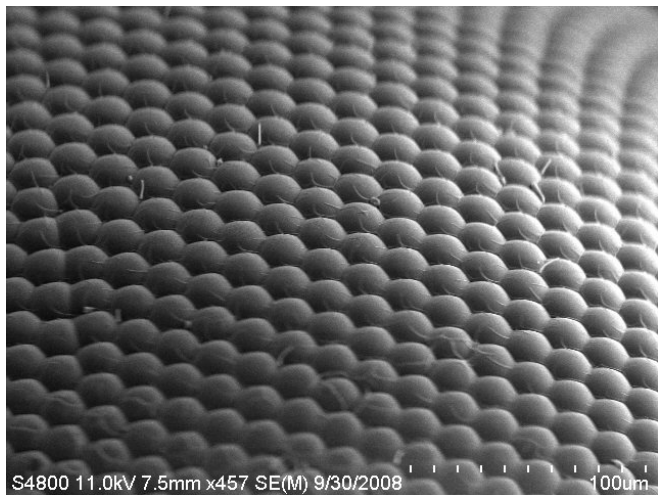


# Scanning electron microscopy (SEM)



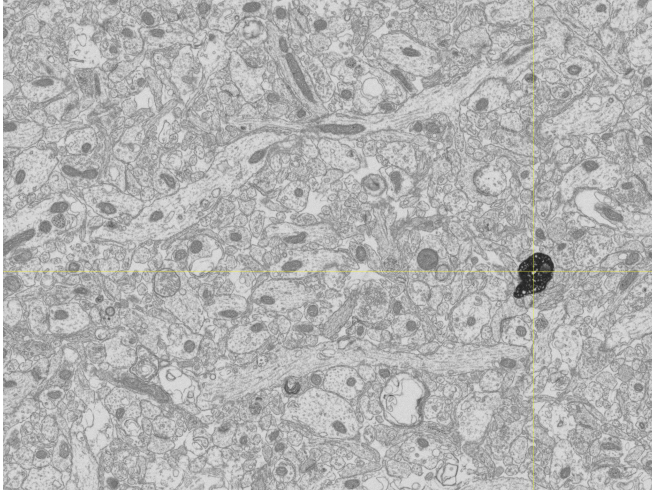
- ▶ Excellent resolution (a few nm)
- ▶ Needs vacuum. Preparation — gold coating, osmium staining, cryofixation.

## SEM example



Fly eye

## FIB example



- ▶ Focused ion beam for slice cutting. True 3D

# Microscopy — types & trends

- ▶ Electron microscopy
  - ▶ Electron transmission microscopy
- ▶ Confocal microscopy — reject out-of-focus light, scanning
- ▶ Two-photon microscopy — long energy fluorescence, reduced phototoxicity, localization of excitation
- ▶ Contrast enhancing techniques
- ▶ Fluorescence microscopy
- ▶ CCD cameras
  - ▶ supercooled
  - ▶ superresolution
- ▶ Moveable specimen tray
  - ▶ Auto-focussing
  - ▶ Automated acquisition, mosaicking

# Microscopy

- ▶ Advantages
  - ▶ High-spatial resolution
  - ▶ Colour and texture information
  - ▶ Affordable (optical microscopy)
  - ▶ Proven technique – large body of experts available
- ▶ Disadvantages
  - ▶ Difficulties of in-vivo observations
  - ▶ Inherently 2D
  - ▶ Missing large-scale perspective