

fMR Processing(2)

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Today goal:

Group analysis of functional activity during Visual task in six subjects using SPM12

- Looking for average activity in group of six measured subjects
 - Is there any common activation? Are the subject's activations on the same place in the brain
1. Do the processing of fMR data for subjects zk84 to zk89 following the pipeline
 2. Do the group statistics and display the results

fMR processing (1. exercise)

Pipeline

Conversion of data:	DICOM -> NIFTI	✓ (done; *.nii suffix)
Slice Timing: temporal correction	(TR vs. HRF)	✗ (skipping this step)

1. **Realignment** of functional data - spatial correction of data in time
2. **Smoothing** with Gauss filter kernel
3. **Model specification + Review**
4. **Estimate Model**
5. **Results + Display**

fMR processing (2. exercise)

Pipeline

Conversion of data:

DICOM -> NIFTI

✓ (done; *.nii suffix)

Slice Timing: temporal correction

(TR vs. HRF)

✗ (skipping this step)

1. **Realignment** of functional data - spatial correction of data in time
2. **Coregister** – T1w image coregister to mean fMR data
3. **Segment** – structural T1w segmentation (WhiteMatter, GrayMatter, CSF)
4. **Normalise** – functional and structural data to a standard MNI space
5. **Smoothing** functional data with Gauss filter kernel
6. **Model specification + Review**
7. **Estimate Model**
8. **Results + Display**

DO IT FOR EACH SUBJECT zk84 – zk89

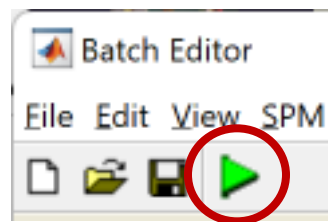
1. Realign data

MENU: Realign (Estimate & Reslice)

BATCH EDITOR:

a) **Data – Session ...** select all fMR files (*MF*.nii*)

b) **Run batch**



Output:

- Resliced files (prefix „r*.nii“)
- „mean*.nii“ file (mean of all fMR files)
- Realignment parameters file „rp_*.txt“

2. Coregister functional and structural data

MENU: Coregister (Estimate)

BATCH EDITOR:

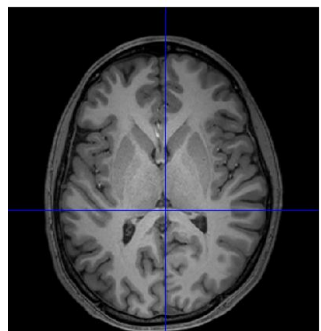
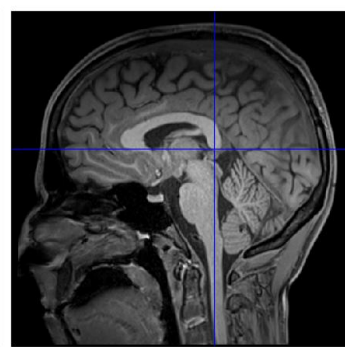
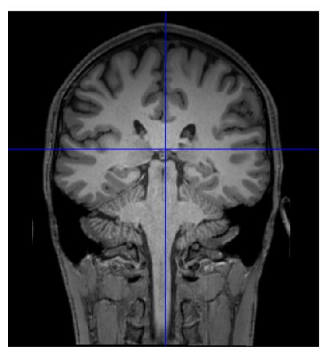
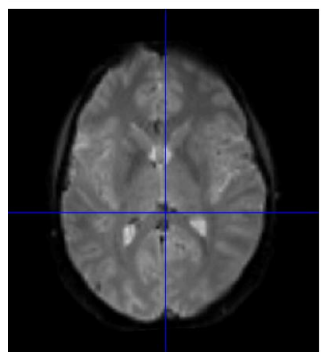
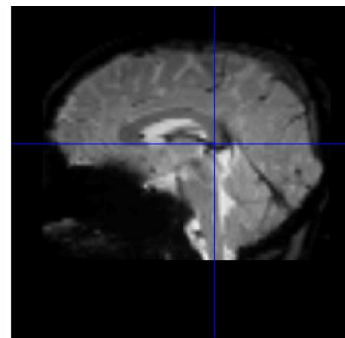
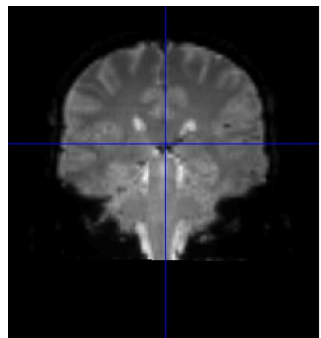
- a) Reference Image ... select meanMF file (*meanMF*.nii*)
- b) Source Image ... select structural T1 data (*zkXX_T1.nii*)
- c) Run batch

- d) Check Reg ... meanMF and zkXX_T1

Output:

- Coregistered zkXX_T1.nii
- No additional files

This step is necessary if fMR and structural data are from different scanners or data were measured in different time



3. Segment structural data

MENU: Segment

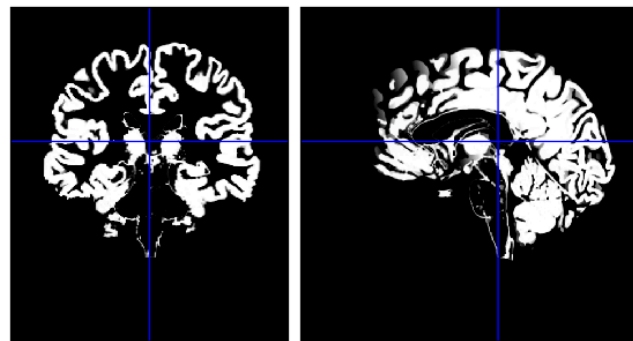
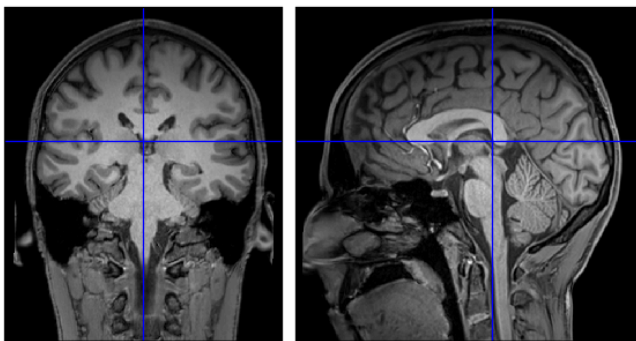
BATCH EDITOR:

- a) **Volumes** ... select structural T1 data (*zkXX_T1.nii*)
- b) **Deformation fields** (last parameter)... *Forward* (transform from functional to structural)
- c) All next parameters as default!
- d) **Run batch** (could be time consuming little bit ☺)
- e) **Display or Check Reg**
output files

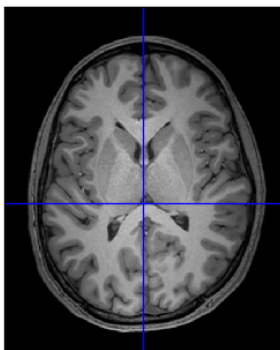
Output:

- c1zkXX_T1.nii (gray matter probability)
- c2zkXX_T1.nii (white matter prob.)
- c3zkXX_T1.nii (CSF prob.)
- y_zkXX_T1.nii (deformation field)
- c4*,c5*zkXX_T1.nii (non brain tissue)

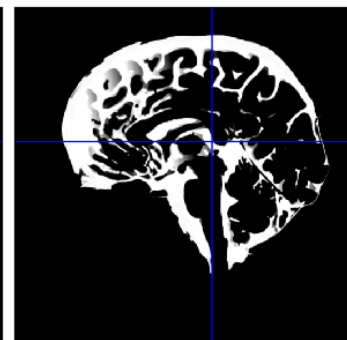
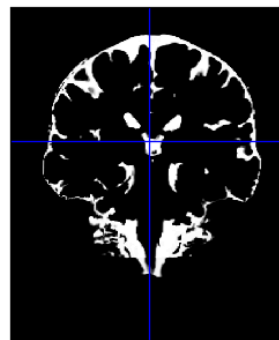
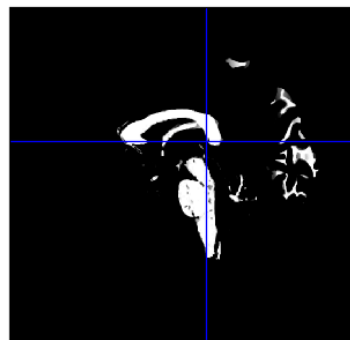
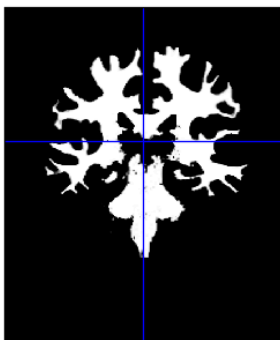
zkXX_T1.nii



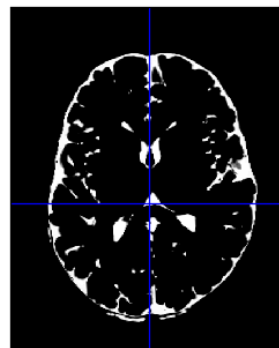
c1*.nii = gray matter



c2*.nii = white matter



c3*.nii = cerebro-spinal fluid



4a. Normalise functional data

MENU: Normalise (Write)

BATCH EDITOR:

- a) **Data – Subject – Deformation Field ...** select Forward deformation Field (*y_zkXX_T1.nii*)
- b) **Images to Write ...** all realigned functional images (*rMF*.nii*)
- c) **Voxel sizes ...** [2 2 2]
- d) **Run batch**

- e) **Display**
output files

Output:

- wrMF*.nii (functional data in standard ***MNI152 space***)

4b. Normalise structural data

MENU: Normalise (Write)

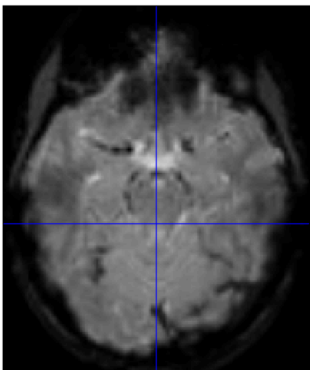
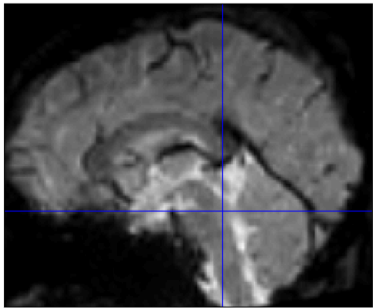
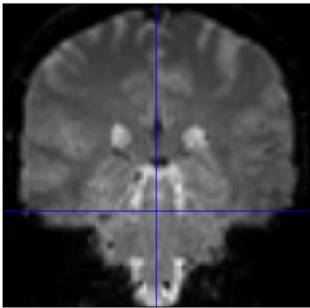
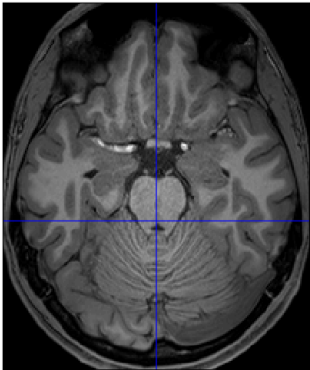
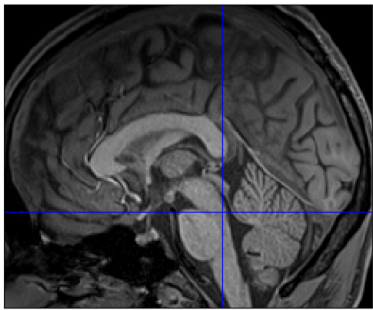
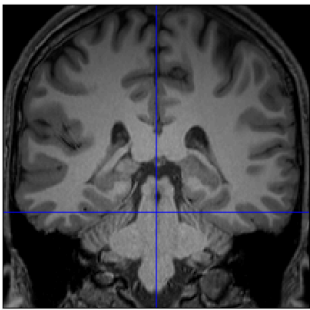
BATCH EDITOR:

- a) **Data – Subject – Deformation Field ...** select Forward deformation Field (*y_zkXX_T1.nii*)
- b) **Images to Write ...** select structural T1 (*zkXX_T1.nii*)
- c) **Voxel sizes ...** [0.75 0.75 0.75]
- d) **Run batch**

- e) **Display**
output files

Output:

- *wzkXX_T1.nii* (structural data in standard ***MNI152 space***)

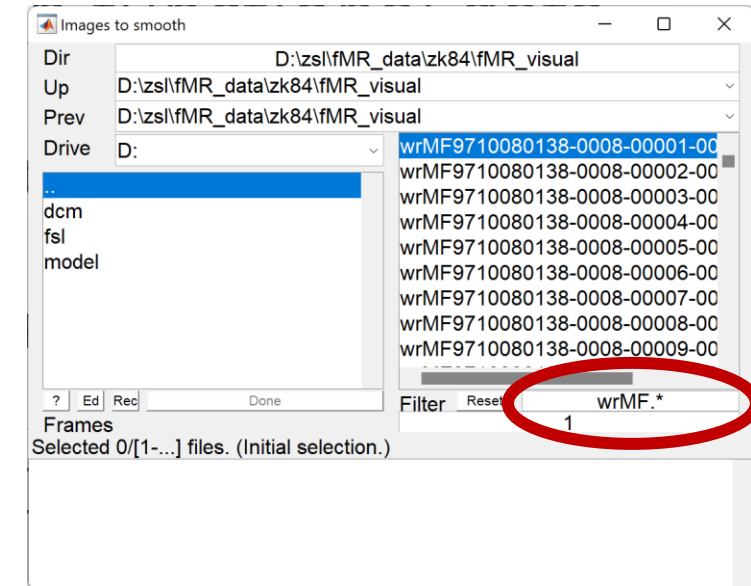


5. Smooth functional data (realigned and normalised)

MENU: Smooth

BATCH EDITOR:

- Images to smooth ...** select all *wrMF*.nii* files
- FWHM ... [8 8 8]** (FWHM of the Gaussian smoothing kernel in mm)
- Run batch**



Output:

- Smoothed functional files (all with prefix „**swr*.nii**“)

6. Model specification

MENU: Specify 1st-level

BATCH EDITOR:

- a) **Directory** ... select (pre-created) folder to store model and statistics results („*model*“)
- b) **Units for design** ... Scans
- c) **Interscan interval** ... 2 (equals to TR)
- d) **Data & Design**
 - a) **Scans** ... select swr*.nii data
 - b) **Conditions**
 - a) **Condition**
 - a) **Name** ... name of processed paradigm, for example VISUAL
 - b) **Onset** ... vector of scans where the activations begins
 - c) **Durations** ... duration of activation (in scans)
- e) **Run batch**

Output:

- **SPM.mat** file (GLM model)

6. Model specification

1. Condition **NEUROTRACKER**

- Name ... NEUROTRACKER
- Onset ... [11 36 61 86 111]
- Durations ... 10

2. Condition **2-BACK**

- Name ... 2BACK
- Onset ... [11 36 61 86]
- Durations ... 15

3. Condition **VISUAL**

- Name ... VISUAL
- Onset ... [16 46 76 106]
- Durations ... 15

7. Model estimation

MENU: Estimate

BATCH EDITOR:

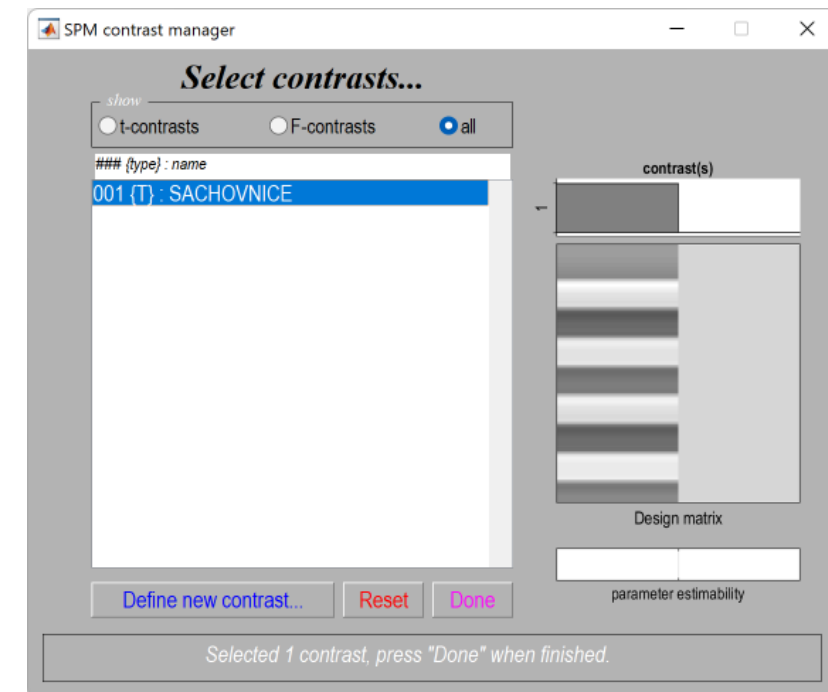
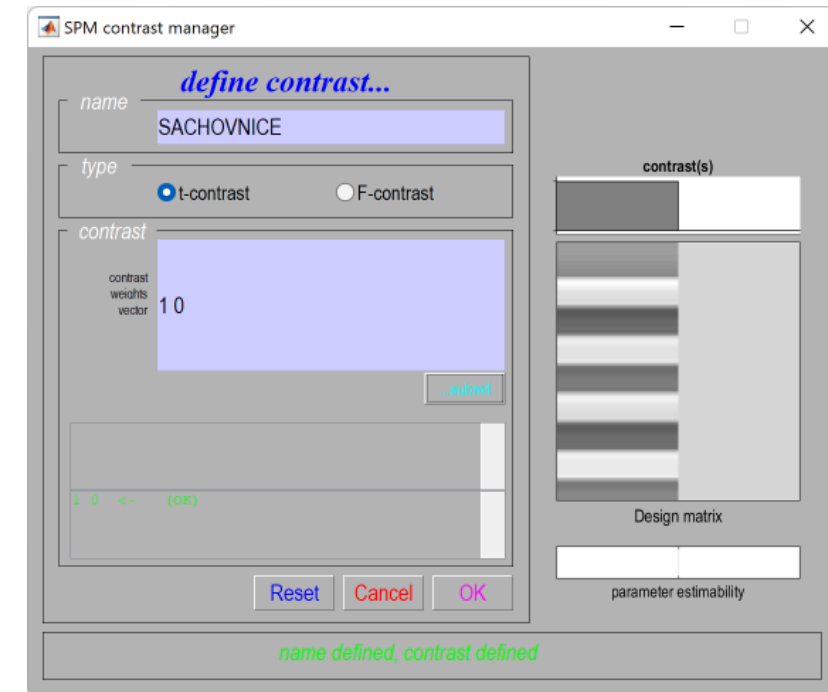
- a) Select SPM.mat ... select *SPM.mat*
- b) Run batch

8. Results (define contrast)

MENU: Results
Select SPM.mat

SPM contrast manager:

- Define new contrast
Name: SACHOVNICE (NEUROTRACKER / 2BACK)
type: t-contrast
contrast: 1 0 ...submit
- OK
- Done, Done



8. Results

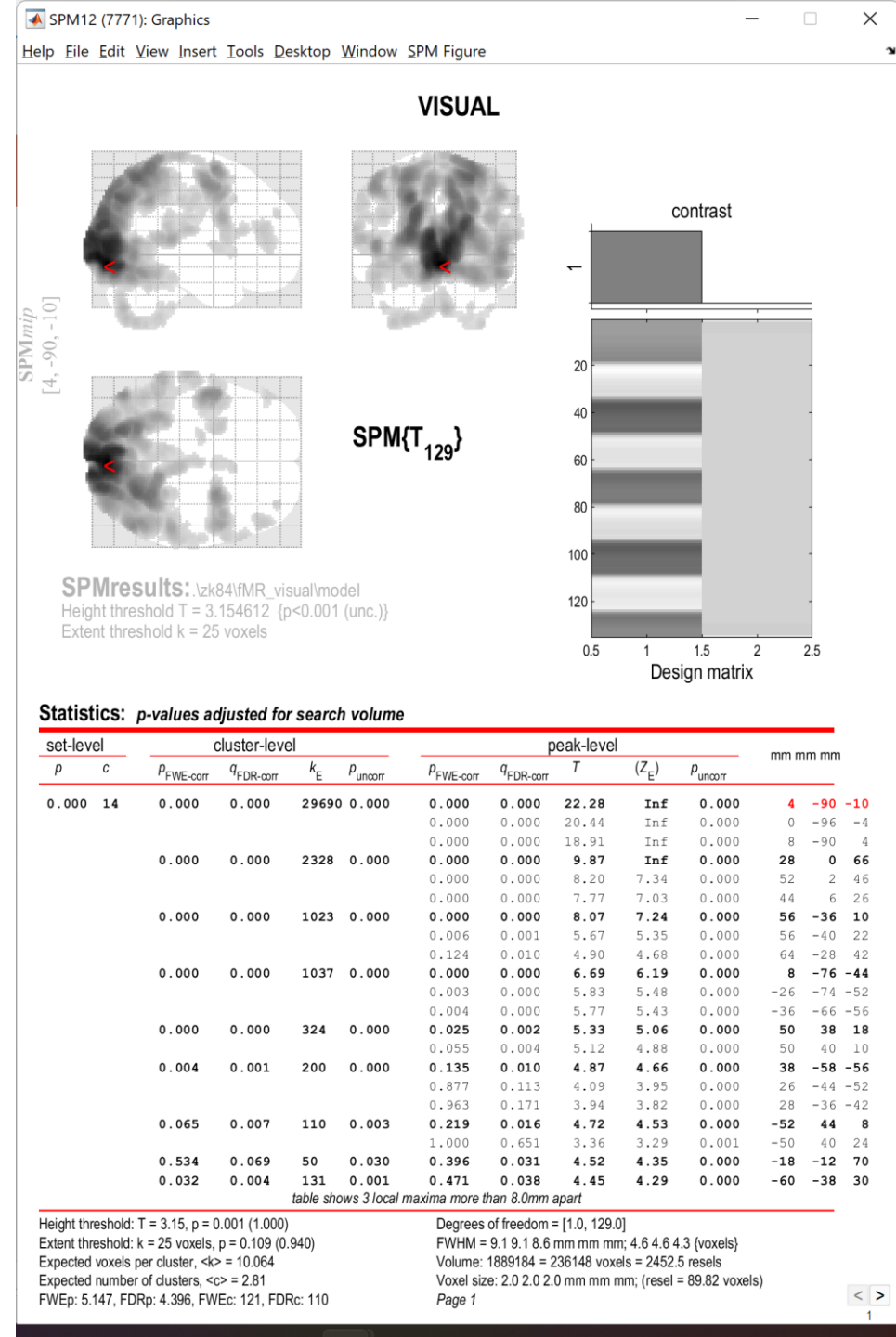
RESULTS:

apply masking ... none

P value adjustment to control ... none

Threshold {T or p value} ... 0.001

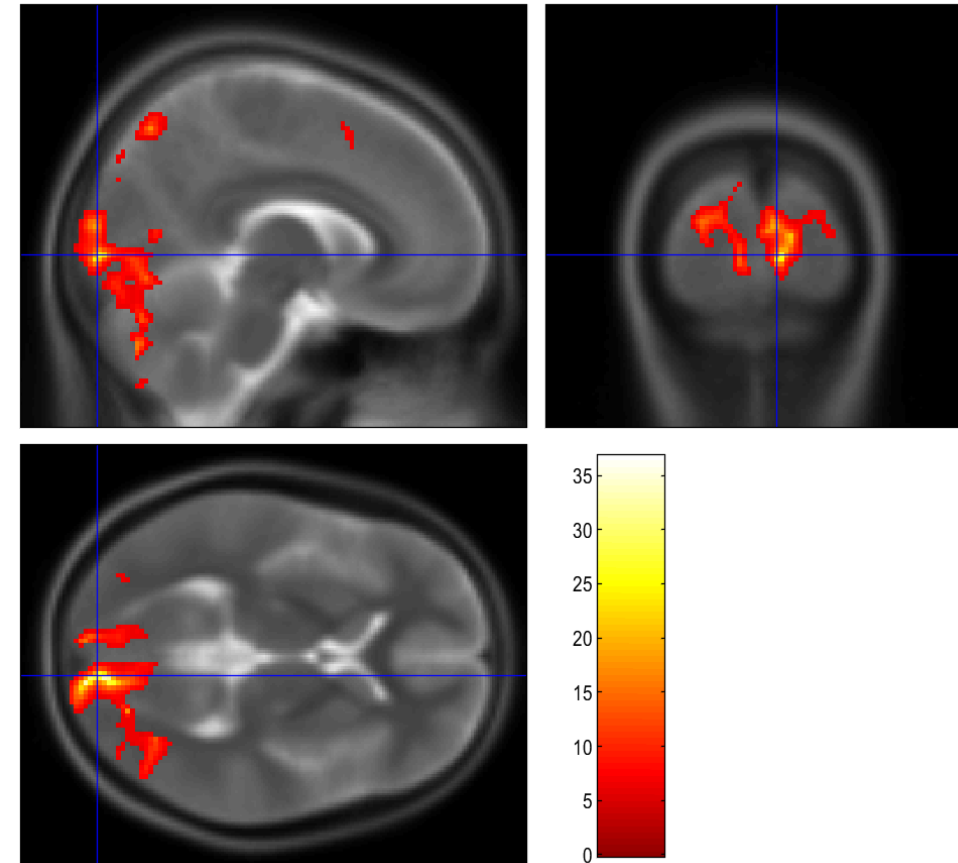
& extent threshold {voxels} ... 25



Display Results

Results can be displayed with **overlay** command
(in results window, Slices, Section, Montage)

As a background image use the MNI template
images from the
`%SPM12%/canonical/avg*.nii`



... do the preview pipeline for subjects

zk84

zk85

zk86

zk87

zk88

zk89

File structure for each subject

- **zkXX**

- **fMR_2back**
- **fMR_neurotracker**
- **fMR_visual**

- *MF*.nii* (135 origin files)
- *rMF*.nii* (135 realigned files)
- *rp_MF*.txt* 1 realignment parameters txt-file
- *meanMF*.nii* 1 mean function file
- *wrMF*.nii* (135 realigned and normalised files)
- *swrMF*.nii* (135 realigned, normalised and smoothed files)
- **model**
 - *beta_0001.nii* beta values of GLM model, for first predictor (predictor of activations)
 - *beta_0002.nii* beta values of GLM model, for second predictor (constant predictor)
 - *con_0001.nii* contrast file for first predictor **IMPORTANT FOR GROUP ANALYSIS!!!**
 - *mask.nii* binary mask file, created automatically
 - *ResMS.nii* residuals file, errors of the GLM model
 - *RPV.nii* residuals per voxel file
 - *SPM.mat* mat-file, where GLM model is stored
 - *spmT_0001.nii* t-statistics T-values

- **t1**

- *c1*.nii* 1 gray matter probability file
- *c2*.nii* 1 white matter probability file
- *c3*.nii* 1 cerebro-spinal fluid probability file
- *y_*.nii* 1 forward deformation field (from structural to MNI space)
- *w*.nii* 1 normalised structural data

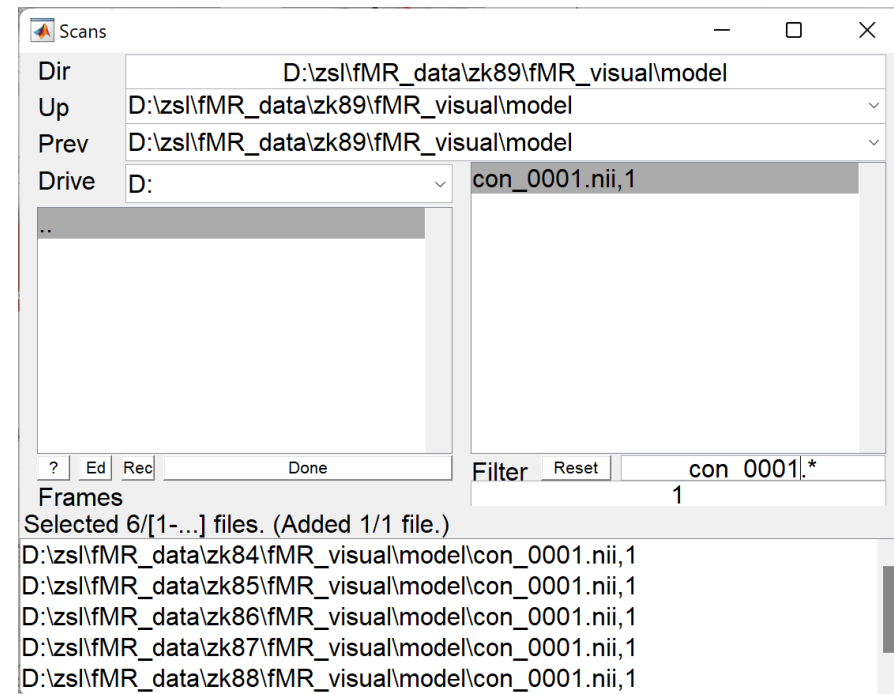
2nd-level
(GROUP ANALYSIS)

9. Model specification (group mean)

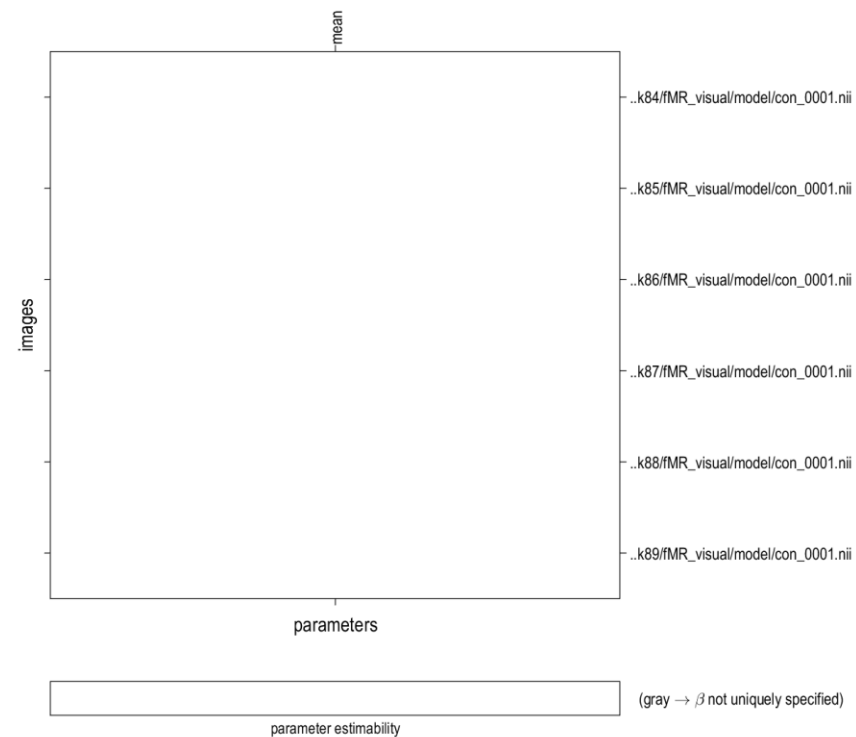
MENU: Specify 2nd-level

BATCH EDITOR:

- Directory ...** select (pre-created) folder (for example „*group_mean*“)
- Design – One-sample t-test – Scans ...** select all *con_0001.nii* files from model folder of each subject (6x *con_0001.nii*)
- Run batch**



Statistical analysis: Design



Design description...

Design : One sample t-test
Global calculation : omit
Grand mean scaling : <no grand Mean scaling>
Global normalisation : <no global normalisation>
Parameters : 1 condition, +0 covariate, +0 block, +0 nuisance
1 total, having 1 degrees of freedom
leaving 5 degrees of freedom from 6 images

10. Model estimation (group mean)

MENU: Estimate

BATCH EDITOR:

- a) Select SPM.mat ... select *SPM.mat*
- b) Run batch

11. Results – define contrast (group mean)

MENU: Results

Select SPM.mat

SPM contrast manager:

a) Define new contrast

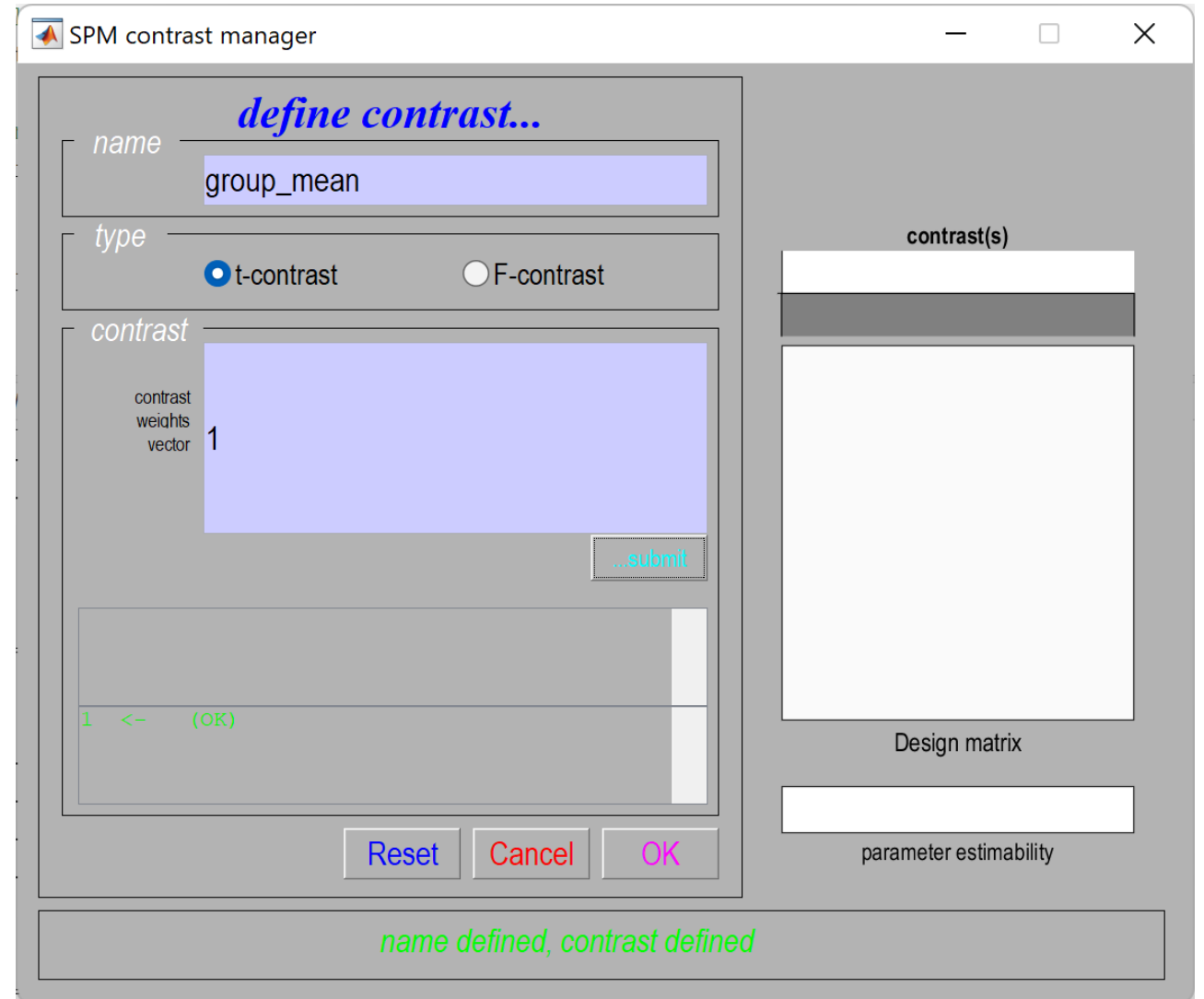
Name: *GROUP_MEAN*

type: *t-contrast*

contrast: *1* ...submit

b) OK

c) Done



11. Results (group mean)

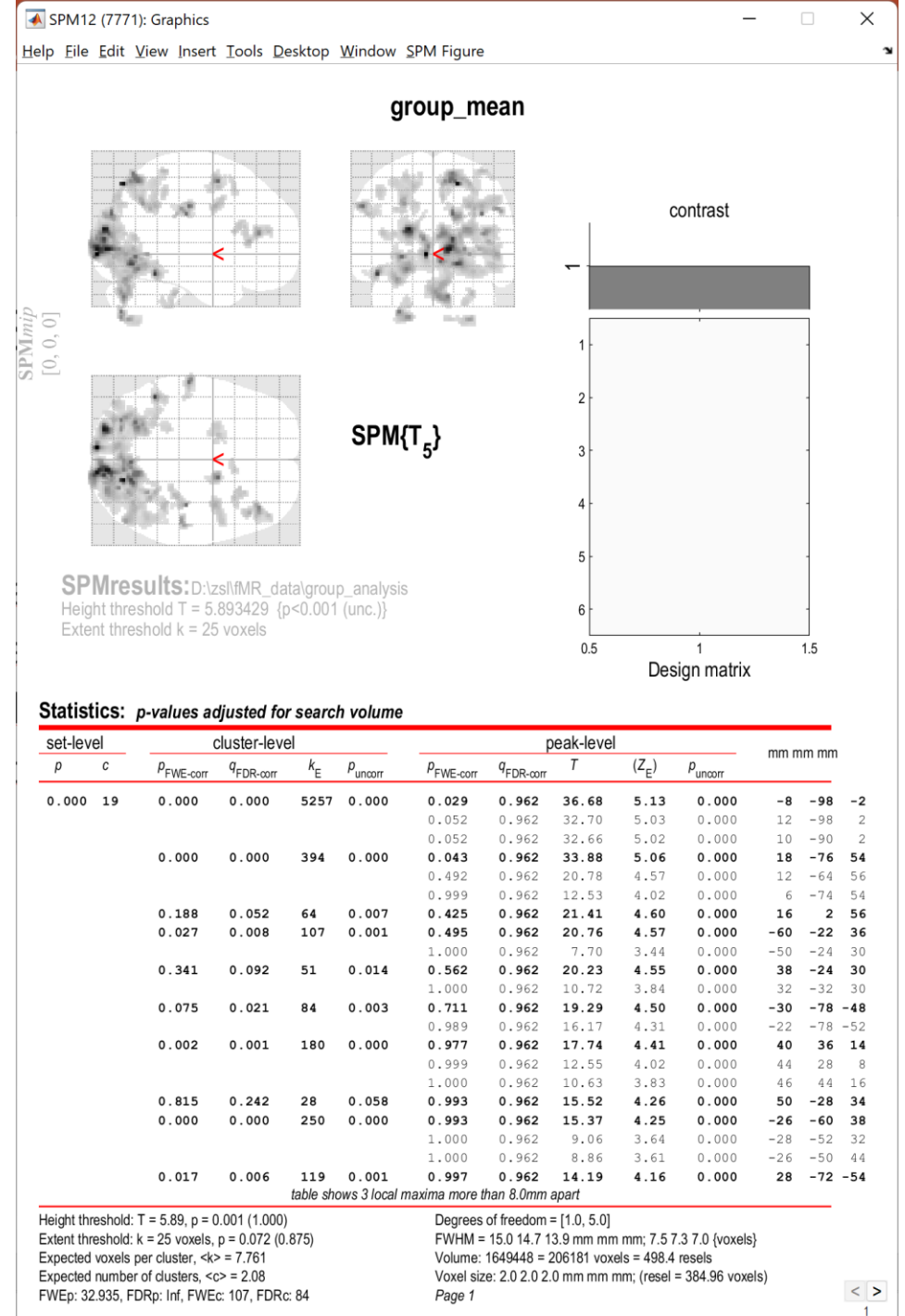
RESULTS:

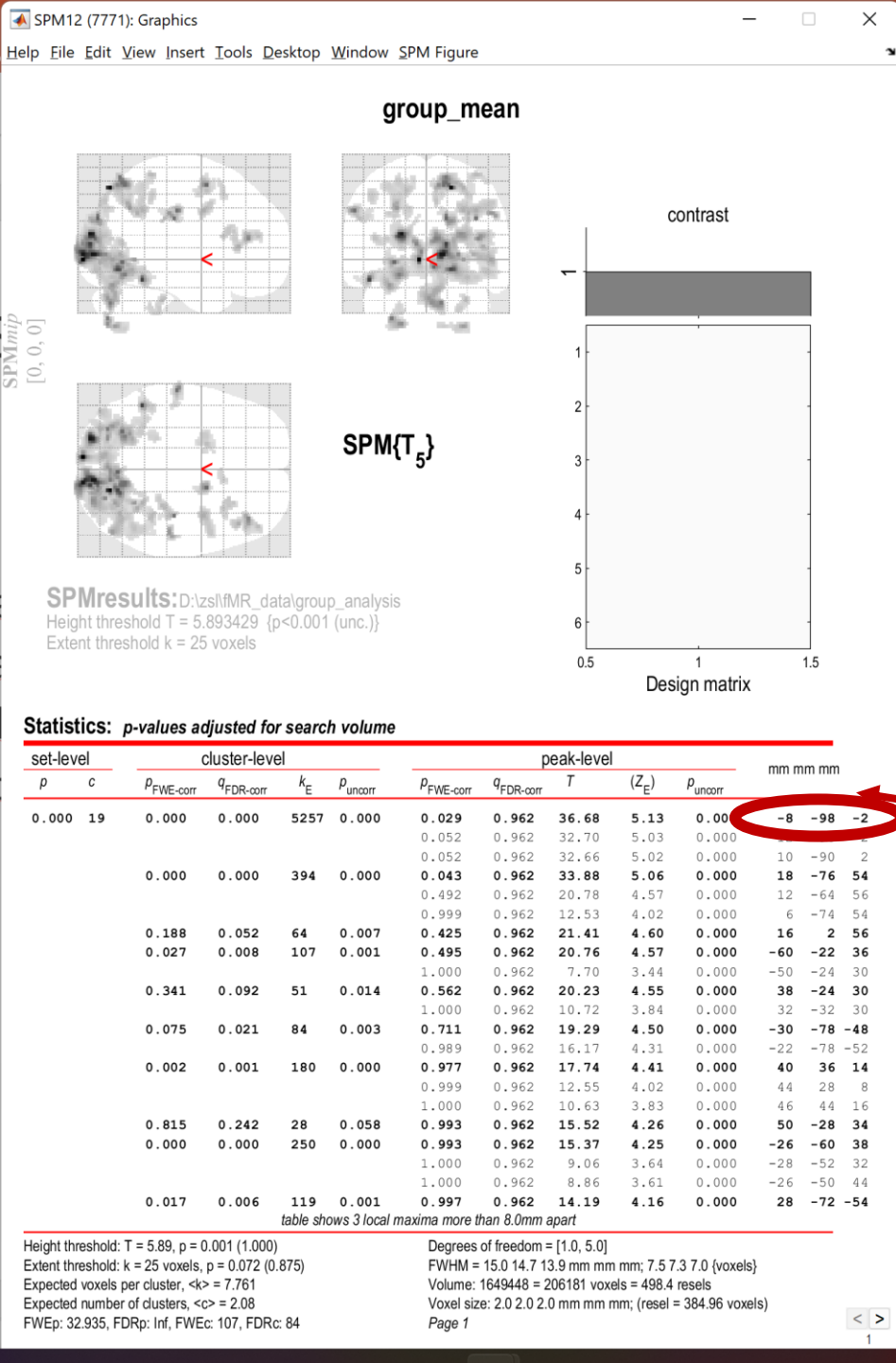
apply masking ... none

P value adjustment to control ... none

Threshold {T or p value} ... 0.001

& extent threshold {voxels} ... 25





Coordinates of maximal activation in MNI space

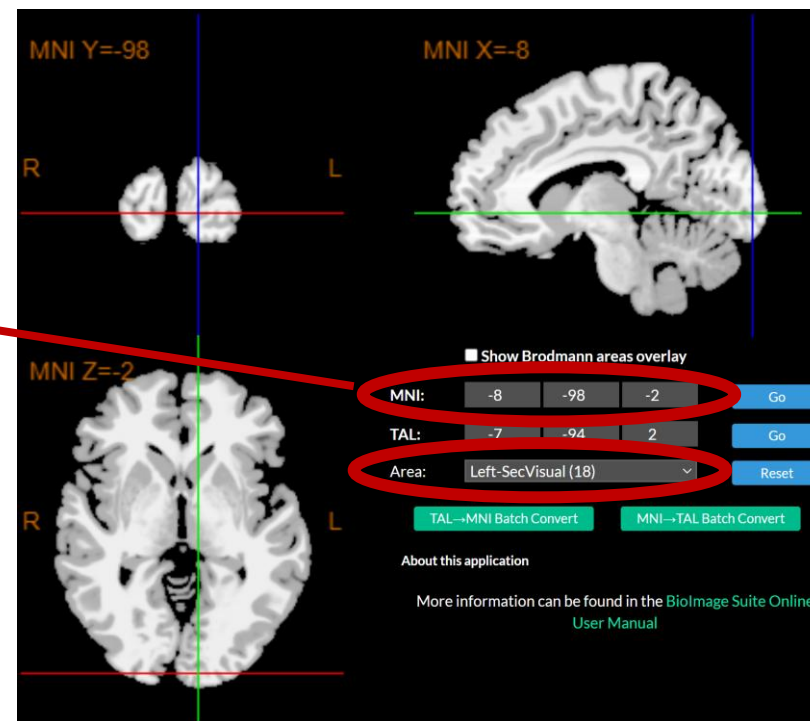
Use

<https://bioimagesuiteweb.github.io/webapp/mni2tal.html>

To find the brain area, which is activated:

List of Brodmann areas:

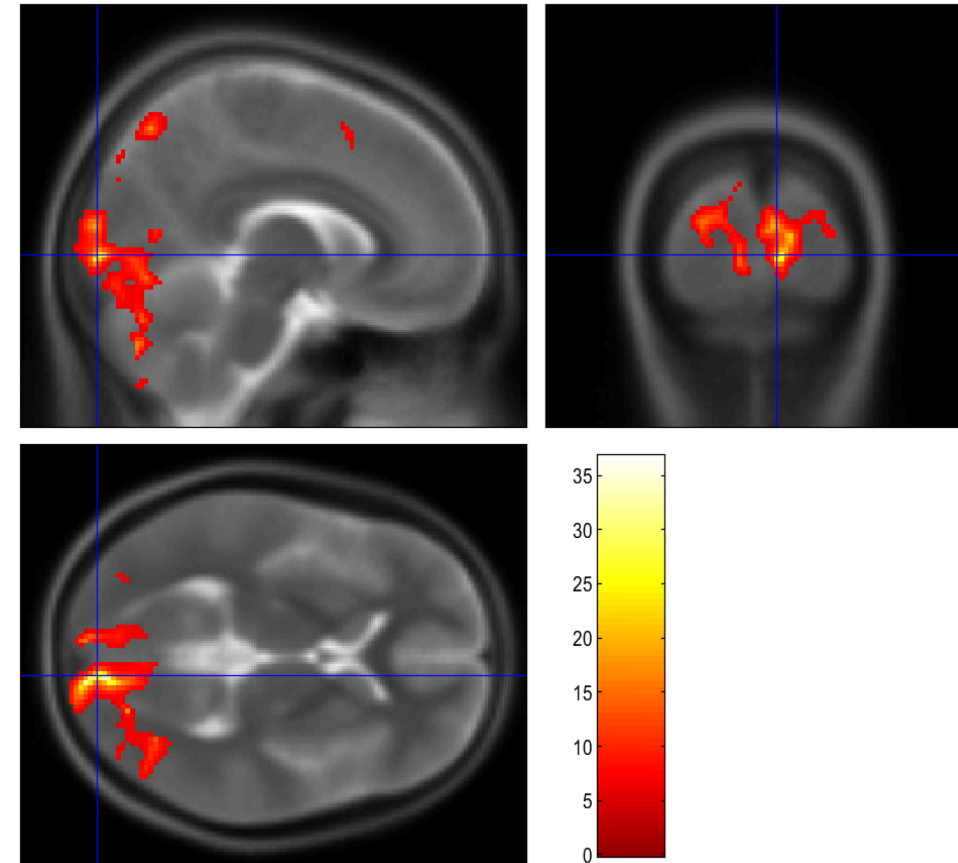
https://en.wikipedia.org/wiki/Brodmann_area



Display Results

Results can be displayed with **overlay** command
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As a background image use the MNI template
images from the
`%SPM12%/canonical/avg*.nii`



Homework

- Choose one of the measured paradigm (neurotracker OR 2back OR visual) and do the processing in all six subjects following the pipeline from the 2nd exercise
- Do the group analysis following the pipeline from the 2nd exercise
- Describe and Display results of the 2nd-level analysis, find areas with statistically significant activation (Brodmann areas)

Hint: batch editor is programable and could be helpful with the repeating commands